Lab on a Chip

TECHNICAL INNOVATION

Dual-mode hydrodynamic railing and arraying of microparticles for multi-stage signal detection in continuous flow biochemical microprocessors

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Continuous flow particulate-based microfluidic processors are in critical demand for emerging applications in chemistry and biology, such as point-of-care molecular diagnostics. Challenges remain, however, for accomplishing biochemical assays in which microparticle immobilization is desired or required during intermediate stages of fluidic reaction processes. Here we present a dual-mode microfluidic reactor that functions autonomously under continuous flow conditions to: (i) execute multi-stage particulate-based fluidic mixing routines, and (ii) array select numbers of microparticles during each reaction stage (e.g., for optical detection). We employ this methodology to detect the inflammatory cytokine, interferon-gamma (IFN-γ), via a six-stage aptamer-based sandwich assay.

Introduction

Particulate-based microfluidic technologies offer significant scaling-induced advantages for biochemical applications, such as pharmacological screening, molecular detection, and quantitative cellular diagnostics.1,2 In particular, bead-based microfluidic systems benefit from high surface-to-volume ratios, rapid reaction kinetics, low reagent volumes, and the ability to functionalize microbeads with surface modifications, including molecular probes capable of detecting a wide range of chemicals and biomolecules.1-5 Consequently, researchers have focused on developing bead-based microfluidic platforms for chemical and biological assays (e.g., immunoassays); however, challenges stemming from the serial fluidic loading requirements associated with such assays have limited the versatility of current systems.5-7 Specifically, biochemical assays primarily utilize multi-stage fluidic processes in which discrete reagents and/or washes are sequentially loaded, resulting in microfluidic systems that typically require external observation and/or regulation during device operation.9-10

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This issue presents additional challenges for fluidic assays that necessitate microbead immobilization (e.g., for visualization and/or fluorescence signal detection) during intermediate steps over the course of such multi-stage processes. Thus, methods to fully automate multi-stage fluidic mixing procedures, while enabling targeted microparticle immobilization, could vastly improve the efficacy of microfluidic biochemical reactors.

Recently, researchers have focused on developing microfluidic techniques to passively transport microparticles into discrete, parallel flow streams under continuous and constant input flow conditions.11-15 Previously, we presented a microfluidic system that utilized microposts arrayed at an angle with respect to the direction of fluid flow to hydrodynamically guide suspended microbeads and living cells into distinct, adjacent flow streams.16 Although we employed this technique to passively accomplish molecular synthesis processes with up to 18 fluidic stages using microbead substrates, immobilization of microparticles was not possible until completion of the full reaction process.16 This is a significant limitation for biochemical assays such as aptamer beacon-based assays that require positive and negative controls of fluorescence intensities prior to reaction completion.3 To overcome this drawback, here we present a dual-mode continuous flow “rail-trap-and-rail” methodology for autonomously executing multi-stage particulate-based microfluidic mixing reactions, while enabling the immobilization of select numbers of microparticles in designated array positions corresponding to each fluidic reaction step (Fig. 1). We utilize this technique to passively accomplish a six-stage aptamer beacon-based sandwich assay to detect the inflammatory cytokine, interferon-gamma (IFN-γ).
Materials and methods

Microfluidic “rail-trap-and-rail” concept

Fig. 1 includes conceptual illustrations of the dual-mode hydrodynamic methodology, which utilizes square-shaped microposts and microparticle trapping sites that are arrayed at a railing angle ($\alpha_R$) or a larger trapping angle ($\alpha_T$), to prevent or promote particle immobilization, respectively (ESI† Fig. 1). Under continuous input flow conditions, the arrayed microposts (of approximately the same size as the target microparticles) passively guide suspended microparticles into discrete, parallel flow streams (without altering the direction of the inputted fluid flow) and toward the trapping area (Fig. 1a). The suspended microparticles passively immobilize in the designated trapping sites, which diverts fluid flow from the occupied traps to the remaining vacant trapping positions. This process facilitates the transport and immobilization of subsequent microparticles into the remaining vacant trapping sites until the trapping area is filled (Fig. 1b). Thereafter, additional suspended microparticles are guided past the previously immobilized particles and into the next micropost array railing area to be transported into a subsequent adjacent fluidic stream (Fig. 1c). This process can be repeated continuously as desired with additional fluidic reagents and/or washes loaded in parallel to customize the microfluidic rail-trap-and-rail system for diverse multi-stage fluidic processes that demand select microparticle immobilization corresponding to intermediate reaction steps.

Continuous flow detection of interferon-gamma (IFN-$\gamma$) via an aptamer beacon-based sandwich assay

To detect the inflammatory cytokine, IFN-$\gamma$, we applied the microfluidic rail-trap-and-rail methodology to execute a six-step aptamer beacon-based sandwich assay under continuous input flow conditions. Fig. 2 includes conceptual illustrations...
of the system architecture and reaction process. The aptamer beacon used in this study was designed previously by Tuleuova et al., and consists of two complementary single-stranded DNA sequences: a fluorescent aptamer (FA), and a quencher (Q) (ESI† Table 1). Six suspensions/solutions were continuously loaded in parallel: (i) a suspension of microbeads functionalized with an avidin-based biological linker, (ii) a solution of biotinylated FAs, (iii) a wash solution of phosphate-buffered saline (PBS), (iv) a solution of Qs, (v) a second solution of PBS, and (vi) a solution of IFN-γ (Fig. 2a). Initially, the functionalized microbeads are transported into the FA solution, which promotes the binding of biotinylated FAs to the microbead substrates via biotin-avidin interactions (Fig. 2b–i–ii). This results in detectable fluorescence on the surface of the microbeads (Fig. 2a–inset). After passing through the PBS wash solution, mixing reactions with the Q solution promote binding of the Qs to complementary FAs, thereby restricting the fluorescence intensity (Fig. 2a; 2b–iii). Following a second wash, additional microbeads are guided into the IFN-γ solution to promote the displacement of Qs by IFN-γ, which enhances the fluorescence response (Fig. 2a; 2b–iv). The fluorescence signals of arrayed microbeads can be detected for each fluidic stage of the assay process (Fig. 2a–inset).

Experimental

Microdevices were fabricated via a previously reported one-mask soft lithography process (ESI† Fig. 2). The devices were designed for 15 μm-in-diameter streptavidin-coated polystyrene microbeads (SVP-150-4, Spherotech, Inc., Lake Forest, IL). The beads were functionalized with an additional biotin-avidin pair to enhance the fluorescence intensities.16 Due to the polydispersity of the microbeads, the devices included microchannel heights of 18 μm, with 5 μm gaps between the microposts (15 × 15 μm²) and traps. The channel lengths for microfluidic mixing were designed as described previously.16 Fabrication results for microfluidic rail-trap-and-rail systems with αα = 1° and αα = 7.5° are shown in ESI† Fig. 3.

The microdevices were pre-treated with Tween 20 (10% in PBS, Fisher Scientific, Pittsburgh, PA).16 We used the Fluenta MAESFLO system to regulate the flow rates (approximately 3 μL min⁻¹) of the six input fluids: (i) microbead suspension (30 beads μL⁻¹), (ii) FA solution (100 μM), (iii) Q solution (100 μM), (iv) IFN-γ solution (10 μM; R&D Systems, Inc, Minneapolis, MN), and (v, vi) two PBS wash solutions (#14287072, Invitrogen Corp., Carlsbad, CA). The full bead-based experimental process was accomplished within 10 minutes. Fluorescence intensities of immobilized microbeads were quantified using ImageJ (NIH, Bethesda, MD). Experimental fluorescence results are presented in the text as mean ± s.e.m.

Results and discussion

Experimental characterization of the effects of the trapping angle (αα) on “rail-trap-and-rail” performance

Experiments were performed using testing systems with αα = 5°, 7.5°, and 10° (while αα was held constant at 1°) in order to examine the effects of αα on device performance. Previously reported equations for quantifying the efficiencies associated with microbead arraying15 and railing16 were used to characterize device performance (ESI† Fig. 4; eqn 1–3). For the αα = 5° testing systems, microbeads often bypassed the trapping areas without being arrayed during the loading process, with primarily

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Fig. 3 Experimental results for microbead dynamics. (a–c) Quantified results for: (a) loading efficiency, (b) trapping efficiency, and (c) railing failure rate (ESI† eqn 1–3) versus varying αα. (d–f) Sequential micrographs of microbeads (15 μm in diameter): (d) railing from the bead suspension into the FA solution, (e) arraying in the designated trapping sites, and (f) railing from the FA solution to the wash solution. Scale bars = 200 μm; αα = 1°; αα = 7.5°. ESI† movie 1 includes video of this process.
Continuous flow bead-based microfluidic detection of IFN-γ

The microfluidic reactor for detecting IFN-γ included raling and trapping areas with $\alpha_r = 5^\circ$ and $\alpha_r = 7.5^\circ$, respectively. Experimental device runs revealed that the microfluidic rail-trap-and-rail methodology successfully transported suspended microbeads into the distinct, parallel reagents and wash solutions, while trapping select numbers of microbeads in the array positions as designed (ESI† movie 1). For example, Fig. 3d–f shows sequential micrographs of microbeads being passively: (d) transported from the bead suspension (white) to the FA solution (cyan), (e) arrayed in the FA solution trapping area, and then (f) guided from the FA solution to the PBS wash solution (white). After microbeads were immobilized in the designated trapping areas (Fig. 4a–e), Relative Fluorescence Intensities (RFIs) were quantified via ESI† eqn 4. The signal intensities exhibited minor variation with time (ESI† Fig. 5). Each of the solutions produced statistically discernable RFIs (Fig. 4f). After mixing with the FA solution, the microbeads exhibited the highest average RFIs of 100% ± 1.3%. Mixing with the Q solution was found to significantly reduce the bead-based fluorescence response ($p < 0.0001$), revealing an average RFI of 0% ± 2.4%. After mixing with the IFN-γ, the average RFI increased significantly to 17.8% ± 1.8% ($p < 0.0001$), which was also discernible from the average RFI corresponding to the second PBS wash solution (10.6% ± 2.2%; $p < 0.05$) (Fig. 4f). These results demonstrate an aptamer-based detection sensitivity of 10 μM of IFN-γ for our assay reaction process.

Conclusions

Continuous flow methods for accomplishing multi-stage microfluidic mixing routines with controlled microparticle arraying are critical to the advancement of diverse chemical and biological applications. Here we presented and demonstrated a dual-mode hydrodynamic methodology by executing a six-stage aptamer beacon-based sandwich assay on microbead substrates under continuous flow conditions. Targeted microbead arraying enabled fluorescence quantification for every phase of the multi-stage process. Here, multi-step microfluidic reactions and analyses were performed to detect the cytokine, IFN-γ; however, microbeads can be functionalized with a variety of molecular probes, and thus, the presented technique could be adapted to accomplish a wide range of multi-step biochemical reaction processes. Additionally, the system in this work included microbeads and microfeatures of approximately the same size, which suggests that this technique could be scaled up or down to handle particles of various sizes. Thus, the presented rail-trap-and-rail methodology could greatly extend the efficacy of particulate-based microfluidic reactors.

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